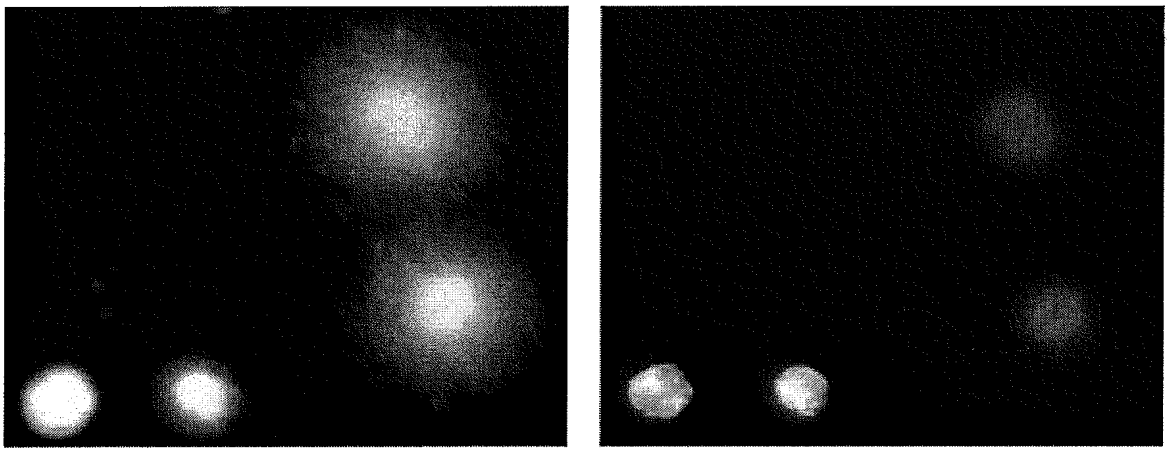

Figure 1. *Sperm cells processed according to the technique of the invention and with Klenow-end labelling for labelling breakages. Only those with a small or no halo, as shown with DAPI staining (blue) (picture in the left, both in the lower left side) have breakage labelling (right, yellow).*

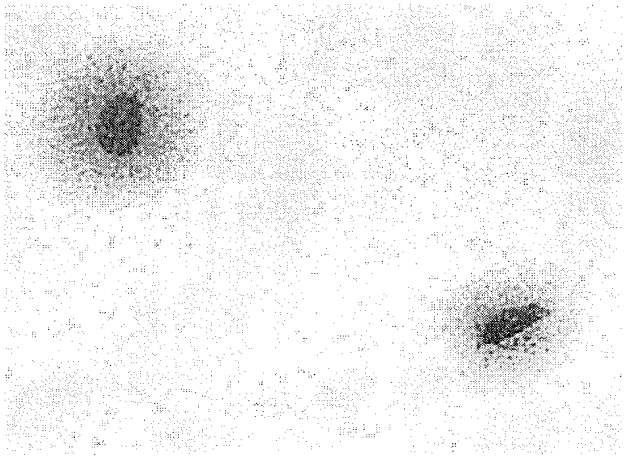

Figure 2. *Solution of lysis with SDS. As shown, the tails do not persist, so it cannot be distinguished if the nucleoid is from a sperm cell or from another cell type present in the sample. In addition, the halos hardly stain with dyes for bright field. This makes imprecise to estimate the limits of the peripheral edge of the halo, i.e., its actual size, so application for patient samples is not possible.*

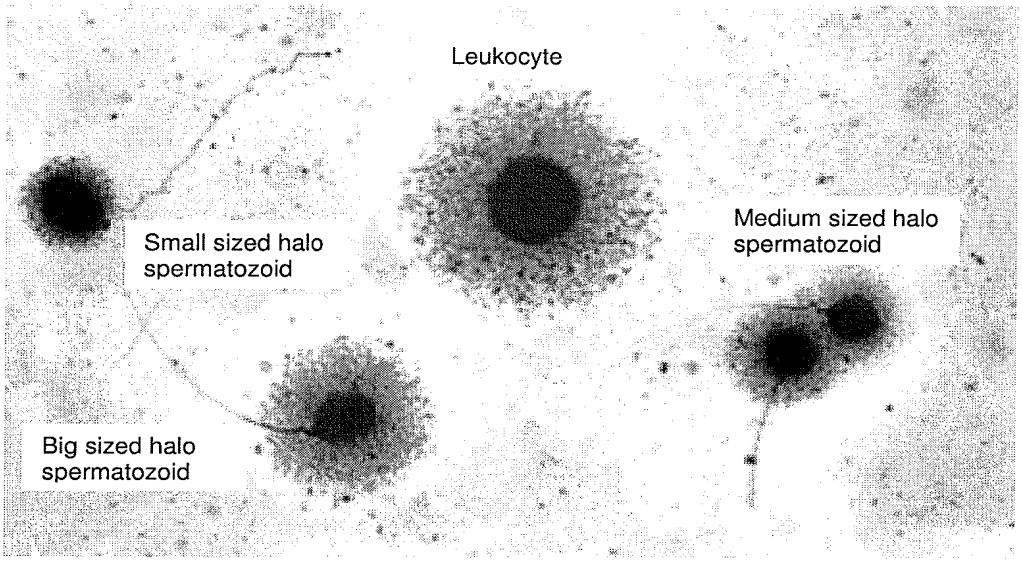

Figure 3. *Solution of lysis with Triton X-100. As shown, the tails are preserved and can be stained with dyes for bright field (Wright staining). Chromatin of the halos is also preserved, so it can be well stained with Wright, and the halo edge can be fully viewed and its size accurately established.*

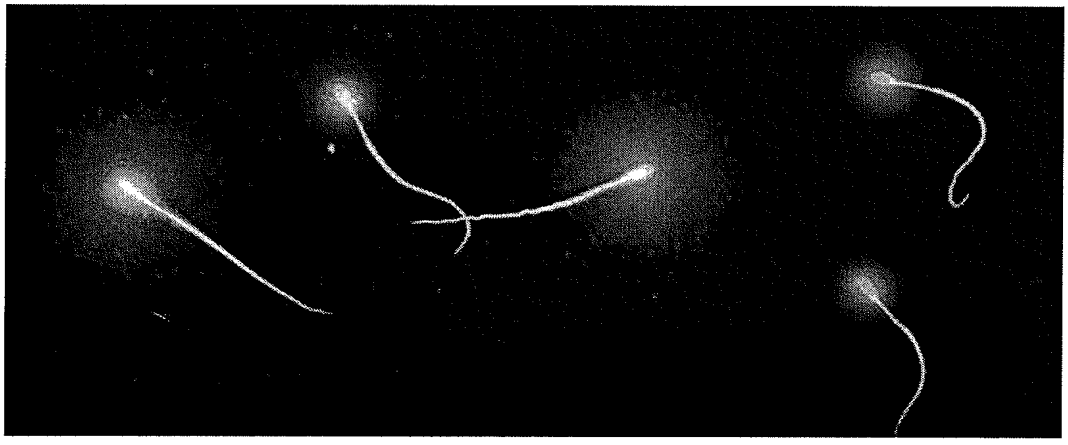

Figure 4. *Horse sperm cells. View of the halos of chromatin dispersion and stability of the flagellum after using a single lysis with triton X-100, in the absence of SDS.*